

# Separation of Empty and Full Engineered Adeno-Associated Virus Capsids Using a Weak Anion Exchanger (Abstract 1038) Varun Gejji\*, Garrett Garrido\*, Erick Reyes-Ramirez\*, Oliver Valle\*, Hari Acharya\* \*Capsida Biotherapeutics, Thousand Oaks, CA 91320

## Introduction

Gene therapy has shown promising results to treat genetic disorders. In this space, recombinant Adeno-associated virus (rAAV) using engineered capsids has gained a lot of attention due to their ability to target specific tissues and improved transduction efficiencies. Capsida has successfully developed unique capsids that cross brain blood barrier (BBB) while de-targeting other organs[1]. As the need to manufacture such highly specific engineered capsids grows, several unit operation process parameters need to be optimized to suffice the growing demand. One of the key consideration is separating full capsids containing the gene of interest (GOI) from empties. Currently, full and empty capsid separation often relies on ultracentrifugation, however, scaling ultracentrifugation is infeasible. In this work, we report successful enrichment of full capsids using anion exchange chromatography (AEX).

# **Methods and Materials**

Engineered AAV capsids (rCap) were first captured using affinity chromatography. The affinity elution pool was buffer exchanged into a low salt solution at a higher (basic) pH, and AEX screening work began using a two-step approach. Initially, different resins (3 strong AEX and 2 weak AEX) were screened and binding strength assessed using SDS-PAGE and Western blot. Based on this initial binding assessment, resins were selected. Elution conditions (different salts, pH conditions) were then screened on 1 mL columns. Elution fractions were analyzed using absorbance ratios, ddPCR, and ELISA based assays. AEX work was performed with three different rCAPs.



## **Results and Discussion**



Figure 2. Initial screening of different AEX resins with multiple novel capsids

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## **PRESENTED ON TUESDAY**

• Chen et al, AAV Gene Therapy Corrects Neurological Phenotypes with Clinically Relevant Doses in a Mouse Model of STXBP1-Related Developmental and Epileptic Encephalopathy (Oral Abs 38)

Figure 1. Downstream Process Flow Overview • Strong AEX resins bound 3 engineered capsids at high pH conditions, however a significant portion of the bound capsids were washed off when the pH was lowered by 0.5 or more units

• One of the weak anion exchangers was protonated at the higher loading pH and failed to bind any of the engineered capsids



- Full and empty capsids co-eluted at low conductivity during gradient elution • Shallow wash with increasing salt concentration removed some empty capsids, however co-elution of full capsids was observed
- Based on absorbance, full enrichment of ~ 1.5x was achieved



- Breakthrough was observed (rCap-4) with strong anion exchange resin at lower pH. The capsid population in the flow-through was mostly empty capsids. However, gradient elution did not further enhance full/empty separation
- Shifting the pH for the load lowered the breakthrough. Shallow wash with increasing salt concentration removed some empty capsids, however the two capsid populations co-eluted at low conductivities
- Based on absorbance, full enrichment of ~2.3x was achieved



- Breakthrough was significantly higher with weak anion exchange resin at high pH. Gradient elution did not improve separation of the two capsid populations.
- Changing the operating pH helped enrich full capsids
- Based on absorbance, the highest enrichment of ~2.7x was achieved

### PRESENTED ON WEDNESDAY

- Knoll et al, CAP-002: Systemic AAV Gene Therapy with Next Generation Capsids for Treatment of STXBP1 Encephalopathy (Abs 504) Shi et al, Directed Evolution of AAV2 • Morales et al, Characterization of engineered AAV capsids from different HEK293 cell culture fractions, crude lysate versus cell pellet material (Abs 529)
- Volpe et al, Alternative Plasmid Designs Including Two Plasmid Transfection Systems for Improved Production and Packaging of Engineered AAV Capsids (Abs 530)







- No breakthrough was observed (rCap-5) with strong anion exchange resin at lower pH
- where empty capsids eluted followed by full capsids

- Based on absorbance, full enrichment was ~2.3x



- were selectively eluted and full capsids were enriched ~2.7x from the starting pool

- performed with the selected resin(s).
- conditions because of unique modifications
- under flow-through mode

### **PRESENTED ON THURSDAY**

Libraries Yields Capsids with Improved Performance in the Central Nervous System and Cross-species Translatability (Abs 992)

### PRESENTED ON FRIDAY

• McDowell et al, Systemic AAV Gene Therapy with CNS-Targeted Engineered Capsids Achieves Significant GCase Activity Increases in the Primate Brain to Support the Potential Treatment of GBA-PD (Oral Abs 274)

• Under a conductivity gradient, full and empty capsids initially co-eluted then a transition was observed

• Performing multiple shallow elution steps helped with selectively desorbing empty capsids

• Two step elution was developed which helped in enriching the full capsids significantly

• With resin D, no capsids were observed in the flowthrough. Using a conductivity shifted wash step, empty capsids

• For Resin C, binding did not occur at the high pH as the resin was completely protonated. Adjusting the load condition and lowering the pH allowed enriching the full capsids as the majority of the empty capsids did not bind

## Conclusions

• Using a two-step AEX development approach, conditions for full enrichment of engineered capsids can be identified. Initially, binding strength for different resins and under different pH conditions is assessed using SDS PAGE and Western blot. Subsequent condition screening is

• Engineered capsids may interact differently with the same AEX resin under the same load

• Understanding the unique modifications on engineered capsids, we are able use capsid specific loading conditions, and empty capsids can be removed from the total population

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